

## Direct Analysis of Urinary Opioids and Metabolites by Mixed-Mode $\mu$ Elution SPE Combined with UPLC/MS/MS for Forensic Toxicology

Jonathan P. Danaceau, Erin E. Chambers, and Kenneth J. Fountain  
Waters Corporation, Milford, MA, USA

### APPLICATION BENEFITS

- Analysis of all metabolites without enzymatic hydrolysis
- Comprehensive panel of 26 opiate and opioid analgesic compounds
- Rapid and simple sample preparation
- Linear response for all analytes and metabolites
- Improved linearity, accuracy and precision vs. dilution protocol
- Reduced matrix effects

### WATERS SOLUTIONS

ACQUITY UPLC® System

ACQUITY UPLC BEH,  
2.1 x 100 mm, 1.7  $\mu$ m Column

Oasis® MCX  $\mu$ Elution Plate

Xevo® TQD Mass Spectrometer

MassLynx™ Software

### KEY WORDS

Opiates, opioids, UPLC, toxicology,  
SPE, sample preparation

### INTRODUCTION

The analysis of natural and synthetic opioid drugs continues to be an important aspect of forensic toxicology. In the past, analyses were typically conducted by GC/MS after first subjecting the samples to acid or enzymatic hydrolysis to liberate glucuronide metabolites.<sup>1</sup> With the advent of LC/MS/MS techniques, glucuronide metabolites can now be analyzed directly.<sup>2-5</sup> Direct analyses of glucuronide metabolites can eliminate the risk of false negatives due to incomplete hydrolysis, as enzymatic efficiency can vary greatly depending upon the enzyme used and the drug substrate analyzed.<sup>6</sup>

Urine samples, unlike some other matrices, can be analyzed by “dilute and shoot” methods in which samples are diluted with an internal standard mix and directly injected onto an LC/MS/MS system.<sup>2,4</sup> Disadvantages to this type of technique, however, include the fact that urine contains many matrix components that can interfere with MS signals. In addition, this technique does not allow for any sample concentration. This can potentially affect the quantification of some of the glucuronide metabolites that elute under high aqueous conditions, where desolvation efficiency is reduced, as well as many of the opioid drugs, since many of them do not produce intense MS/MS product fragments.

This application note highlights a method for the analysis of 26 opioid drugs and metabolites by mixed-mode SPE followed by UPLC®/MS/MS. Glucuronide metabolites are directly analyzed, eliminating the need for enzymatic or chemical hydrolysis. Direct comparison demonstrates that mixed-mode SPE has improved linearity, greater accuracy and precision, and fewer matrix effects than a simple dilute and shoot method. Previously confirmed, incurred samples were also analyzed, allowing for additional evaluation of this method.

**EXPERIMENTAL****LC Conditions**

LC system:	ACQUITY UPLC
Column:	BEH C <sub>18</sub> , 2.1 x 100 mm, 1.7 µm, part number 186002352
Column temp.:	30 °C
Injection volume:	10 µL
Flow rate:	0.4 mL/min
Mobile phase A:	0.1% formic acid in MilliQ® water
Mobile phase B:	0.1% formic acid in ACN
Gradient:	Initial conditions were 2% B. The %B was increased to 47.2% over 6.0 min and then returned to 2% over 0.5 min. The system was allowed to reequilibrate for 1.5 min. The entire cycle time was 8.0 min.

**MS Conditions**

MS system:	Xevo TQD Mass Spectrometer
Ionization mode:	ESI <sup>+</sup>
Acquisition mode:	MRM (See Table 1 for transitions)
Capillary voltage:	1 kV
Collision energy (eV):	Optimized for individual compounds (See Table 1)
Cone voltage (V):	Optimized for individual compounds (See Table 1)
Data Management:	All data were acquired and analyzed using MassLynx Software v.4.1

**Materials**

All compounds and internal standards (IS) were purchased from Cerilliant® (Round Rock, TX). Complementary, deuterated internal standards were used for all compounds with the exception of hydromorphone-3-glucuronide, codeine-6-glucuronide, norbuprenorphine-glucuronide, norfentanyl, and buprenorphine-glucuronide. For these compounds, a deuterated IS with the most similar response was chosen as a surrogate.

A combined stock solution of all compounds (10 µg/mL; 2.5 µg/mL for fentanyl and norfentanyl) was prepared in methanol. Working solutions were made daily by preparing high standards and QCs in matrix (urine) and performing serial dilutions to achieve the desired concentrations. Calibrator concentrations ranged from 5 to 500 ng/mL for all analytes with the exception of fentanyl and norfentanyl, which were prepared at 25% of the concentration of the other analytes (1.25 to 125 ng/mL). A combined internal standard stock solution (5 µg/mL; 1.25 µg/mL for fentanyl and norfentanyl) was prepared in methanol. Working IS solutions were prepared daily in MilliQ water at 50 ng/mL.

**Sample Preparation**

Sample preparation consisted of either simple dilution or mixed-mode SPE. For the dilution method, 100 µL of urine was diluted 1:1 with MilliQ water containing internal standards. The samples were vortexed and then loaded into individual wells in the collection plate. For mixed-mode SPE, urine samples (method blanks, standards, QCs and unknowns) were pretreated by adding equal amounts of 4% H<sub>3</sub>PO<sub>4</sub> and a working IS mixture (50 ng/mL) prepared in MilliQ water. Wells in the Oasis MCX µElution 96-well plate (part number 186001830BA) were conditioned with 200 µL MeOH followed by 200 µL MilliQ water. 300 µL of each prepared sample was then added to each well, resulting in a sample load of 100 µL urine. After loading, the wells were washed with another 200 µL water followed by 200 µL MeOH. All samples were then eluted with 2 x 50 µL of 60:40 MeOH/ACN containing 5% of a concentrated NH<sub>4</sub>OH solution (Fisher, 20-22%). After elution, all samples were evaporated under N<sub>2</sub> to dryness (approximately 5 min) and reconstituted with a solution of 98:2 water/ACN containing 0.1% formic acid and 0.1% human plasma. 10 µL was injected onto the LC/MS/MS system.

## RESULTS AND DISCUSSION

The 26 compounds and metabolites screened are listed in Table 1 and constitute a comprehensive panel of natural opiate drugs, semi-synthetic opioids, and synthetic narcotic analgesic compounds. Most all of the compounds are weak bases, with pKa values of approximately 8 to 9. They have a wide range of polarities, with LogP values ranging from -3.48 for morphine-3 $\beta$ -d-glucuronide to 5.00 for methadone, as shown in Table 1; MRM transitions used are also listed there.

Compound	RT	Formula	Molecular Mass	LogP (predicted)	MRM Transitions	Cone Voltage	Coll. Energy
1 Morphine-3 $\beta$ -D-glucuronide	1.21	C <sub>23</sub> H <sub>27</sub> NO <sub>9</sub>	461.17	-3.48	462.1>286.1 462.1>201.1	58 58	30 52
2 Oxymorphone-3 $\beta$ -D-glucuronide	1.21	C <sub>23</sub> H <sub>27</sub> NO <sub>10</sub>	477.16	-	478.1>284.1 478.1>227.1	46 46	28 50
3 Hydromorphone-3 $\beta$ -D-glucuronide	1.34	C <sub>23</sub> H <sub>27</sub> NO <sub>9</sub>	461.17	-	462.1>286.1 462.1>185.1	58 58	28 56
4 Morphine-6 $\beta$ -D-glucuronide	1.47	C <sub>23</sub> H <sub>27</sub> NO <sub>9</sub>	461.17	-2.98	462.2>286.2 462.2>201.2	64 64	38 40
5 Morphine	1.50	C <sub>17</sub> H <sub>19</sub> NO <sub>3</sub>	285.14	0.90	286.2>201.1 286.2>165.1	54 54	28 34
6 Oxymorphone	1.61	C <sub>17</sub> H <sub>19</sub> NO <sub>4</sub>	301.13	0.78	302.1>227.1 302.1>242.1	44 44	28 24
7 Hydromorphone	1.76	C <sub>17</sub> H <sub>19</sub> NO <sub>3</sub>	285.13	1.62	286.2>185.1 286.2>157.1	66 66	32 42
8 Codeine-6 $\beta$ -D-glucuronide	2.00	C <sub>24</sub> H <sub>29</sub> NO <sub>9</sub>	475.18	-2.84	476.2>300.2 476.2>165.2	60 60	36 40
9 Dihydrocodeine	2.07	C <sub>18</sub> H <sub>23</sub> NO <sub>3</sub>	301.17	1.55	302.2>199.1 302.2>128.1	52 52	34 58
10 Codeine	2.14	C <sub>18</sub> H <sub>21</sub> NO <sub>3</sub>	299.15	1.34	300.2>215.2 300.2>165.1	54 54	26 38
11 Oxycodone	2.37	C <sub>18</sub> H <sub>21</sub> NO <sub>4</sub>	315.15	1.03	316.2>256.2 316.2>241.1	44 44	26 26
12 6-Acetylmorphine (6-AM)	2.41	C <sub>19</sub> H <sub>21</sub> NO <sub>4</sub>	327.15	1.31	328.2>165.1 328.2>211.1	60 60	26 36
13 O-desmethyl Tramadol	2.46	C <sub>15</sub> H <sub>23</sub> NO <sub>2</sub>	249.17	1.72	250.2>58.0	26	18
14 Hydrocodone	2.50	C <sub>18</sub> H <sub>21</sub> NO <sub>3</sub>	299.15	1.96	300.2>199.1 300.2>171.0	60 60	30 44
15 Norbuprenorphine-glucuronide	2.83	C <sub>31</sub> H <sub>43</sub> NO <sub>10</sub>	589.29	-	590.3>414.3 590.3>101.0	70 70	34 54
16 Norfentanyl	2.93	C <sub>14</sub> H <sub>20</sub> N <sub>2</sub> O	232.16	1.42	233.2>177.2 233.2>150.1	30 30	14 18
17 Tramadol	3.21	C <sub>16</sub> H <sub>25</sub> NO <sub>2</sub>	263.19	2.45	264.2>58.0	24	16
18 Normeperidine	3.58	C <sub>14</sub> H <sub>19</sub> NO <sub>2</sub>	233.10	2.07	234.1>160.1 234.1>188.2	36 36	12 18
19 Meperidine	3.60	C <sub>15</sub> H <sub>21</sub> NO <sub>2</sub>	247.16	2.46	248.2>174.1 248.2>220.2	48 48	22 20
20 Buprenorphine-glucuronide	3.64	C <sub>35</sub> H <sub>49</sub> NO <sub>10</sub>	643.34	-	644.3>468.3 644.3>187.1	66 66	42 62
21 Norbuprenorphine	3.77	C <sub>25</sub> H <sub>35</sub> NO <sub>4</sub>	413.26	2.30	414.3>101.0 414.3>187.2	66 66	42 34
22 Fentanyl	4.29	C <sub>22</sub> H <sub>28</sub> N <sub>2</sub> O	336.22	3.82	337.2>188.2 337.2>105.1	48 48	22 38
23 Buprenorphine	4.55	C <sub>29</sub> H <sub>41</sub> NO <sub>4</sub>	467.30	3.55	468.3>101.0 468.3>396.3	72 72	40 48
24 EDDP*	4.79	C <sub>20</sub> H <sub>24</sub> N*	278.19	-	278.3>234.2 278.3>249.2	50 50	24 32
25 Propoxyphene	5.18	C <sub>22</sub> H <sub>29</sub> NO <sub>2</sub>	339.30	4.90	340.3>266.2 340.3>143.1	22 22	8 32
26 Methadone	5.25	C <sub>21</sub> H <sub>27</sub> NO	309.20	5.01	310.2>105.0 310.2>223.1	32 32	22 28

Table 1. Chemical properties and MS conditions of test compounds.

## Chromatography

During the initial chromatographic method development, two types of acidic additives (buffers) were evaluated. One was 0.1% formic acid and the second was a combination of 2 mM ammonium acetate with 0.1% formic acid, a mobile phase similar to one used in a related application.<sup>7</sup> No substantial differences in chromatography were seen. However, the analytical sensitivity of several compounds was significantly suppressed when using the combination of ammonium acetate and formic acid. The peak area of all of the glucuronide metabolites and norbuprenorphine were reduced by 60% to 80% compared to those seen with formic acid alone. Thus, the remaining experiments were conducted with the mobile phases containing 0.1% formic acid alone.

A representative chromatogram of all compounds from a 50 ng/mL calibration standard is shown in Figure 1. Peak assignments can be found in Table 1. Using an ACQUITY UPLC BEH C<sub>18</sub>, 2.1 x 100 mm, 1.7 μm Column we were able to analyze all analytes in under 5.5 min with baseline separation between all critical pairs of isomers, such as between morphine-3-glucuronide, morphine-6-glucuronide, and hydromorphone-3-glucuronide (compounds 1, 3, and 4, respectively) and near baseline separation between morphine-6-glucuronide and morphine.

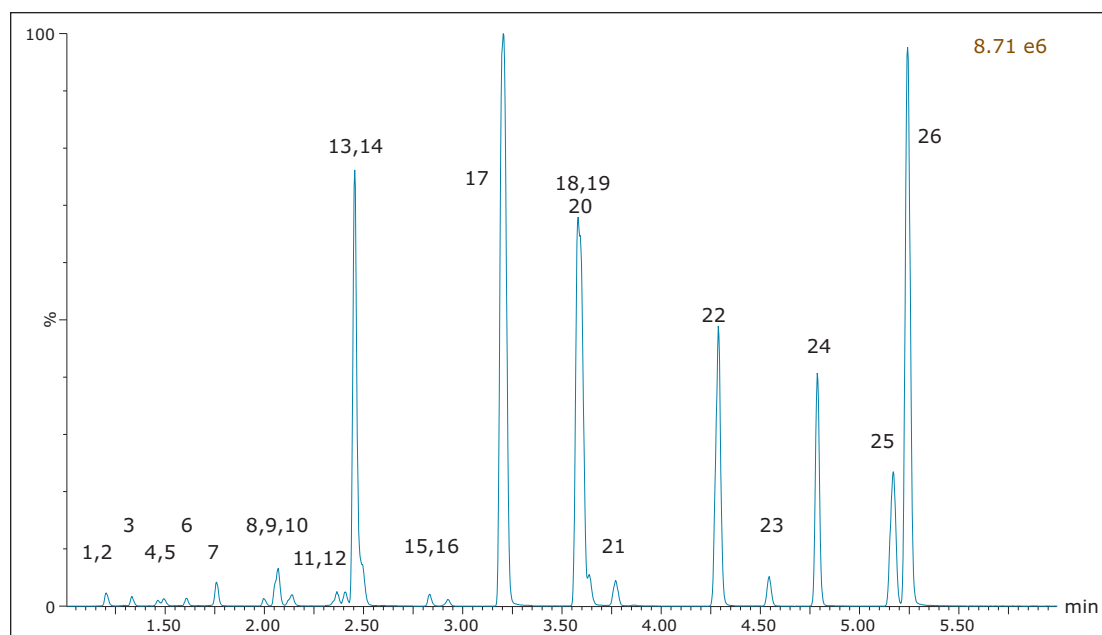


Figure 1. Chromatography of opiate and synthetic analgesic compounds. Peak assignments are listed in Table 1.

## Recovery and Matrix Factors

Both mixed-mode SPE and simple dilution were evaluated as possible sample preparation methods. Sample dilution has the advantages of being very simple, inexpensive, and, in the case of urine samples, compatible with reversed-phase chromatographic conditions. Disadvantages include reduced analytical sensitivity resulting from sample dilution and potential interference from matrix components remaining in the sample. SPE, on the other hand, can reduce potential matrix effects because of its selective nature. In addition, the ability of SPE to concentrate the sample can help improve analytical sensitivity of the assay. For this application, evaporation of the organic eluate and reconstitution in a high aqueous solution (2% ACN) was necessary to prevent solvent effects that otherwise interfered with the chromatography of the glucuronide metabolites. Figure 2 shows the average recovery of all compounds from six different lots of urine using the Oasis MCX  $\mu$ Elution protocol detailed above. With the exception of the four earliest eluting glucuronide metabolites, all compounds demonstrated recoveries of 89% or greater. In addition, when peak areas from extracted 50 ng/mL samples were compared, the areas for the Oasis MCX  $\mu$ Elution protocol ranged from 2.1 to more than six times greater than the dilution protocol. Thus, the ability to concentrate the samples more than made up for the limited recovery seen for a few analytes.

In addition to recovery, matrix factors were evaluated for both protocols. Matrix factors were calculated according to the following equation:

$$\text{Matrix Factor (MF)} = (\text{peak area in the presence of matrix}) / (\text{peak area in the absence of matrix})$$

In the case of SPE, blank urine was subjected to the extraction protocol, and standards (dissolved in methanol) were added to the final eluate. For the solvent standard, the same methanolic standard solution was combined with 50  $\mu$ L of the elution solution. Both groups of samples were then evaporated and reconstituted as previously described. For dilution samples, diluted urine samples spiked with drug standards were compared to samples consisting of the reconstitution solution spiked with drug standards.

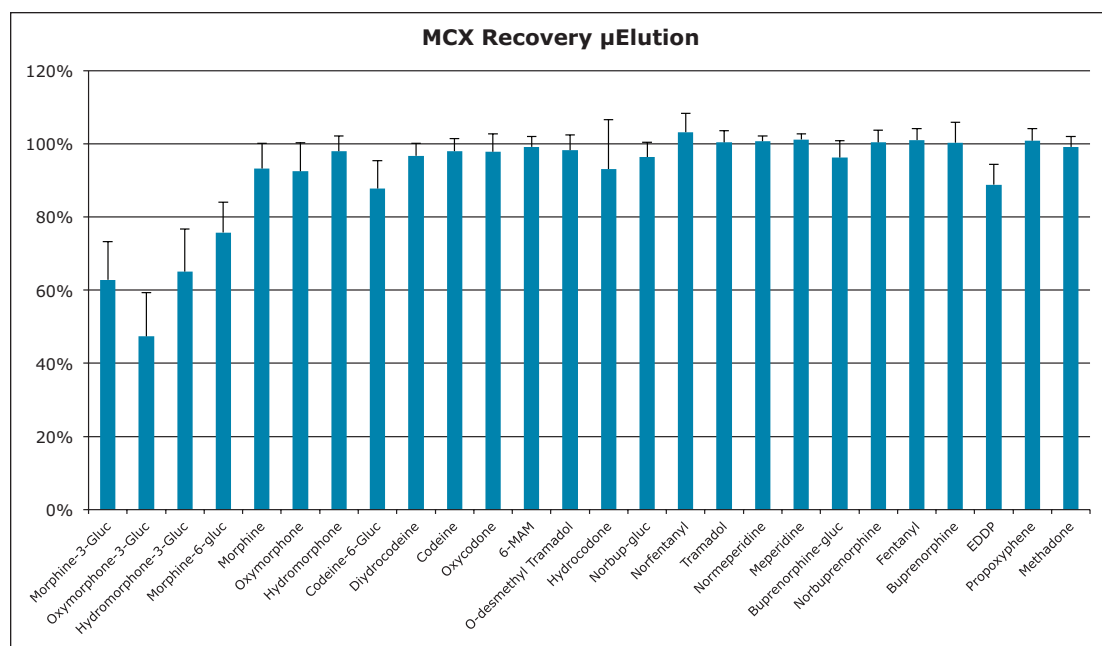


Figure 2. Recovery of opioid compounds from urine using Oasis MCX  $\mu$ Elution Plates. Bars represent the mean recovery from six lots of urine.

Figure 3 shows the results of the matrix factor experiments conducted with six different lots of urine. While both protocols show the trend toward suppression of the earlier eluting compounds, statistical analysis reveals that nearly half of the compounds (12 of 26) demonstrated significantly less matrix interference when the Oasis MCX  $\mu$ Elution protocol was used. The asterisks in the figure indicate those compounds in which matrix factors were significantly different between the two protocols. In every case in which a significant difference was observed, mixed-mode SPE resulted in matrix factors closer to the ideal value of 1 (no matrix effect). In addition, matrix factors were more consistent when using the mixed-mode SPE protocol. With the exception of oxymorphone (17.0%), oxycodone (15.9%), and fentanyl (20.6%), all compounds in the SPE prepared samples had coefficients of variation (CVs) of less than 15.0%. By contrast, only 12 of the compounds prepared by sample dilution had CVs less than 15.0%. Thus, the use of mixed-mode SPE resulted in not only reduced matrix effects, but also resulted in less variability among different lots of urine.

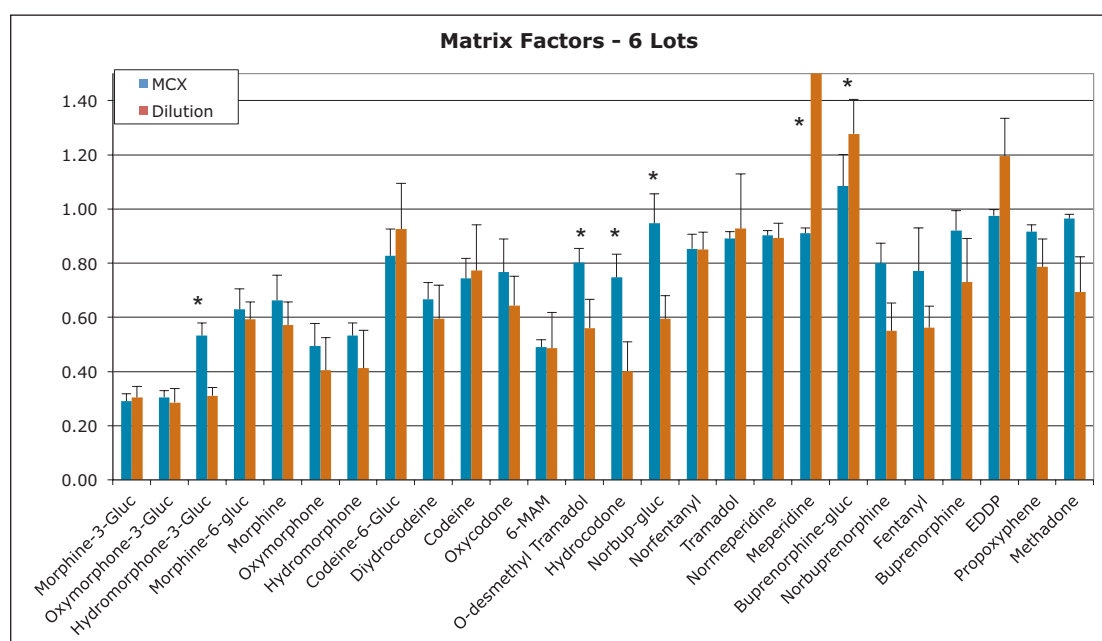


Figure 3. Mean matrix effects of opioid compounds from six lots of urine. Blue bars indicate matrix effects measured from Oasis MCX  $\mu$ Elution Plates. Red bars indicate matrix effects resulting from sample dilution. Asterisks indicate compounds in which the difference between the two protocols was significantly different.

## Linearity

The two sample preparation protocols were also evaluated for linearity and accuracy. Calibration standards were prepared in urine at concentrations ranging from 5 to 500 ng/mL (1.25 to 125 ng/mL for fentanyl and norfentanyl). Quality control samples (N=4) were prepared at four concentrations: 7.5, 75, 250 and 400 ng/mL. These samples were then prepared by either mixed-mode SPE or sample dilution. The mean accuracies and  $R^2$  values for the calibration curves are shown in Tables 2 and 3. For the SPE prepared samples, the means of all calibration points were within 10% of their expected values. The American Association of Clinical Chemistry (AACC) suggests that %CVs be less than 10%, a criterion which is met by all points with the exception of morphine at 10 and 500 ng/mL and morphine-6-glucuronide at 5 ng/mL. All compounds show excellent linearity, with  $R^2$  values of 0.992 or greater.



### Accuracy and Precision

A similar pattern seen in the calibration curves is observed when looking at quality control results for both methods. Table 4 reveals that, with the exception of morphine at 7.5 ng/mL, %CVs for all compounds prepared by mixed-mode SPE fall within the suggested precision requirements of < 10% at all four QC concentrations. With very few exceptions, nearly all accuracy and precision values are less than 10%. In addition, only three QC points show a deviation from expected values of more than 10% and all are within 15%. By contrast, the results for samples prepared by the dilution protocol show that many compounds fail precision (%RSD) requirements, especially at the lower concentration of 7.5 ng/mL, as shown in Table 5, and many values deviate from their expected concentrations by more than 15%, especially at the low QC concentration.

	QC Concentration (ng/mL)											
	7.5			75			250			400		
	Mean	%CV	Bias	Mean	%CV	Bias	Mean	%CV	Bias	Mean	%CV	Bias
Morphine-3-β-d-glucuronide	7.10	8.3%	-5.3%	74.5	5.2%	-0.7%	250.0	2.2%	0.0%	386.3	3.6%	-3.4%
Oxymorphone-3-β-d-glucuronide	7.43	9.7%	-1.0%	76.9	3.0%	2.5%	239.9	4.9%	-4.0%	372.1	3.7%	-7.0%
Hydromorphone-3-β-d-glucuronide	7.98	7.8%	6.3%	76.4	5.8%	1.9%	252.4	2.9%	0.9%	398.1	3.7%	-0.5%
Morphine-6-gluc	8.30	8.7%	10.7%	74.9	6.7%	-0.1%	240.9	5.1%	-3.7%	376.8	4.0%	-5.8%
Morphine	8.15	10.1%	8.7%	75.6	7.7%	0.8%	217.1	5.1%	-13.2%	391.2	4.3%	-2.2%
Oxymorphone	7.85	5.1%	4.7%	73.3	4.2%	-2.3%	243.6	4.7%	-2.6%	385.5	4.5%	-3.6%
Hydromorphone	7.93	1.6%	5.7%	75.7	3.0%	0.9%	247.8	3.7%	-0.9%	388.9	1.2%	-2.8%
Codeine-6-β-d-glucuronide	7.78	4.0%	3.7%	73.6	3.8%	-1.9%	257.3	5.0%	2.9%	421.7	2.6%	5.4%
Dihydrocodeine	7.65	0.8%	2.0%	75.8	1.1%	1.1%	243.8	0.6%	-2.5%	377.9	2.8%	-5.5%
Codeine	7.68	4.7%	2.3%	75.8	0.6%	1.1%	245.2	1.9%	-1.9%	385.4	0.9%	-3.7%
Oxycodone	7.58	5.2%	1.0%	75.5	2.3%	0.7%	244.5	3.4%	-2.2%	378.0	2.8%	-5.5%
6-Acetylmorphone (6-AM)	7.70	5.3%	2.7%	76.2	4.3%	1.6%	245.9	2.3%	-1.7%	391.5	0.7%	-2.1%
O-desmethyl Tramadol	7.83	1.9%	4.3%	75.0	1.3%	0.0%	247.1	0.7%	-1.2%	384.6	0.7%	-3.8%
Hydrocodone	7.60	1.9%	1.3%	74.5	1.3%	-0.7%	244.2	1.6%	-2.3%	381.3	0.9%	-4.7%
Norbuprenorphine-glucuronide	7.80	3.6%	4.0%	76.4	3.1%	1.8%	255.0	3.9%	2.0%	401.9	1.3%	0.5%
Norfentanyl	1.90	0.0%	1.3%	19.4	2.3%	3.3%	62.7	1.2%	0.4%	101.7	2.2%	1.7%
Tramadol	7.60	0.0%	1.3%	76.8	0.3%	2.4%	240.5	0.8%	-3.8%	369.2	0.5%	-7.7%
Normeperidine	7.48	2.0%	-0.3%	75.3	1.6%	0.4%	238.7	1.2%	-4.5%	371.4	1.4%	-7.2%
Meperidine	7.43	0.7%	-1.0%	73.2	0.5%	-2.5%	242.4	2.4%	-3.1%	388.1	1.7%	-3.0%
Buprenorphine-gluc	8.08	2.7%	7.7%	77.8	1.8%	3.7%	267.0	1.6%	6.8%	441.1	1.3%	10.3%
Norbuprenorphine	7.73	1.2%	3.0%	77.7	3.8%	3.6%	246.1	1.5%	-1.6%	377.2	1.0%	-5.7%
Fentanyl	1.90	0.0%	1.3%	19.2	1.1%	2.4%	60.8	1.0%	-2.7%	96.8	1.0%	-3.2%
Buprenorphine	7.55	2.3%	0.7%	77.2	1.9%	2.9%	247.2	1.9%	-1.1%	397.1	1.3%	-0.7%
EDDP+	7.65	1.3%	2.0%	75.0	1.1%	0.0%	243.2	0.9%	-2.7%	387.7	1.1%	-3.1%
Propoxyphene	7.55	0.8%	0.7%	78.4	0.5%	4.5%	243.4	0.9%	-2.6%	378.9	1.9%	-5.3%
Methadone	7.58	0.7%	1.0%	78.2	1.5%	4.3%	246.4	1.0%	-1.4%	386.4	1.2%	-3.4%

■ %RSD > 10% or %deviation >15%

Table 4. Quality control statistics for opioid compounds extracted using Oasis MCX μElution Plates. For each concentration, mean, %CV and % bias are listed (N=4).



	QC Concentration (ng/mL)											
	7.5			75			250			400		
	Mean	%RSD	Bias	Mean	%RSD	Bias	Mean	%RSD	Bias	Mean	%RSD	Bias
Morphine-3-β-d-glucuronide	7.08	10.3%	-5.7%	73.3	6.1%	-2.3%	239.4	2.3%	-4.2%	380.0	6.2%	-5.0%
Oxymorphone-3-β-d-glucuronide	6.85	18.1%	-8.7%	72.9	6.8%	-2.8%	229.7	4.0%	-8.1%	365.9	7.0%	-8.5%
Hydromorphone-3-β-d-glucuronide	7.75	14.5%	3.3%	78.1	4.5%	4.1%	236.5	6.9%	-5.4%	362.7	5.8%	-9.3%
Morphine-6-gluc	7.85	23.1%	4.7%	74.0	17.5%	-1.4%	249.1	9.3%	-0.4%	358.6	3.5%	-10.4%
Morphine	5.28	26.9%	-29.7%	76.0	7.9%	1.3%	267.4	9.4%	7.0%	410.7	16.6%	2.7%
Oxymorphone	8.98	23.3%	19.7%	82.4	9.7%	9.9%	251.9	5.8%	0.7%	360.1	5.4%	-10.0%
Hydromorphone	8.13	14.1%	8.3%	79.3	5.0%	5.7%	251.8	5.1%	0.7%	381.1	3.4%	-4.7%
Codeine-6-β-d-glucuronide	6.45	11.5%	-14.0%	71.6	7.0%	-4.5%	226.7	8.0%	-9.3%	358.6	4.4%	-10.4%
Dihydrocodeine	8.25	9.4%	10.0%	86.1	8.0%	14.8%	244.8	5.6%	-2.1%	387.3	5.3%	-3.2%
Codeine	7.90	10.5%	5.3%	76.5	4.7%	2.0%	236.2	8.0%	-5.5%	366.0	3.9%	-8.5%
Oxycodone	7.53	20.4%	0.3%	79.2	6.8%	5.6%	243.0	3.4%	-2.8%	380.3	3.4%	-4.9%
6-Acetylmorphone (6-AM)	6.50	7.7%	-13.3%	68.3	9.5%	-8.9%	215.6	2.8%	-13.8%	371.6	5.2%	-7.1%
O-desmethyl Tramadol	7.45	3.6%	-0.7%	79.5	4.9%	5.9%	240.2	3.3%	-3.9%	369.0	2.5%	-7.8%
Hydrocodone	6.75	8.2%	-10.0%	71.9	3.6%	-4.2%	227.2	6.4%	-9.1%	341.2	5.8%	-14.7%
Norbuprenorphine-glucuronide	7.25	5.3%	-3.3%	77.1	2.7%	2.8%	234.5	5.0%	-6.2%	350.2	3.0%	-12.4%
Norfentanyl	1.53	11.2%	-18.7%	20.1	3.7%	6.9%	60.3	3.7%	-3.6%	92.1	0.6%	-7.9%
Tramadol	6.53	1.5%	-13.0%	69.8	3.6%	-6.9%	218.1	1.3%	-12.8%	335.5	0.8%	-16.1%
Normeperidine	7.45	4.6%	-0.7%	79.3	5.1%	5.7%	234.6	3.1%	-6.2%	356.8	0.7%	-10.8%
Meperidine	7.33	1.7%	-2.3%	77.4	7.0%	3.2%	236.3	2.1%	-5.5%	367.0	2.7%	-8.2%
Buprenorphine-gluc	4.80	4.5%	-36.0%	65.8	3.6%	-12.3%	211.1	4.9%	-15.6%	327.1	2.1%	-18.2%
Norbuprenorphine	7.15	9.2%	-4.7%	79.6	2.8%	6.2%	242.6	5.6%	-3.0%	364.2	1.7%	-9.0%
Fentanyl	1.75	3.3%	-6.7%	19.5	2.9%	3.9%	60.0	3.9%	-4.1%	91.9	1.4%	-8.2%
Buprenorphine	6.80	6.4%	-9.3%	75.5	3.8%	0.6%	231.1	3.7%	-7.6%	356.6	2.3%	-10.9%
EDDP+	7.45	1.7%	-0.7%	78.3	3.3%	4.4%	239.2	1.0%	-4.3%	365.2	2.1%	-8.7%
Propoxyphene	7.00	8.2%	-6.7%	75.9	2.2%	1.2%	229.7	2.8%	-8.1%	349.9	4.5%	-12.5%
Metadone	6.98	6.0%	-7.0%	75.6	2.5%	0.7%	232.8	3.4%	-6.9%	349.5	4.4%	-12.6%

■ %RSDs > 10% or %deviation > 15%

Table 5. Quality control statistics for opioid compounds prepared using a simple sample dilution protocol. For each concentration, mean, %CV and % bias are listed (N=4).

## Analysis of Incurred Samples

In order to test this method in a real-world context, 32 urine samples (two negative, 30 positive) previously confirmed for opiate compounds were obtained and analyzed by the current method. These samples had been analyzed for 6-MAM (heroin metabolite), codeine, hydrocodone, hydromorphone, morphine, oxycodone, and oxymorphone. Among the differences in analysis was the fact that these samples had been hydrolyzed to release the conjugated metabolites from the glucuronide moieties. Figure 4 compares the results obtained from the current method to those reported from the laboratory that provided the samples for oxycodone and hydrocodone. These two compounds both lack hydroxyl groups at positions three and six. This renders them incapable of undergoing phase two glucuronidation,<sup>6,8</sup> eliminating any discrepancies in the data due to incomplete hydrolysis. These two figures show fairly good correlation when comparing the two methods, with R<sup>2</sup> values of 0.956 and 0.985 for hydrocodone and oxycodone, respectively. With a slope of near 1 (m=0.962), the oxycodone results between the two methods are in good agreement. For hydrocodone, there is a bias towards higher concentrations in the method presented here (m=0.689). This could be due to the influence of two highly concentrated samples with measured concentrations of 6574 and 7032 ng/mL by the current method that had previously reported results of 3750 and 4610 ng/mL, respectively. For the current analysis, these samples were diluted to concentrations within the reported linear range of 5 to 500 ng/mL. It is unknown if the previously reported results represented samples that had been properly diluted or not.

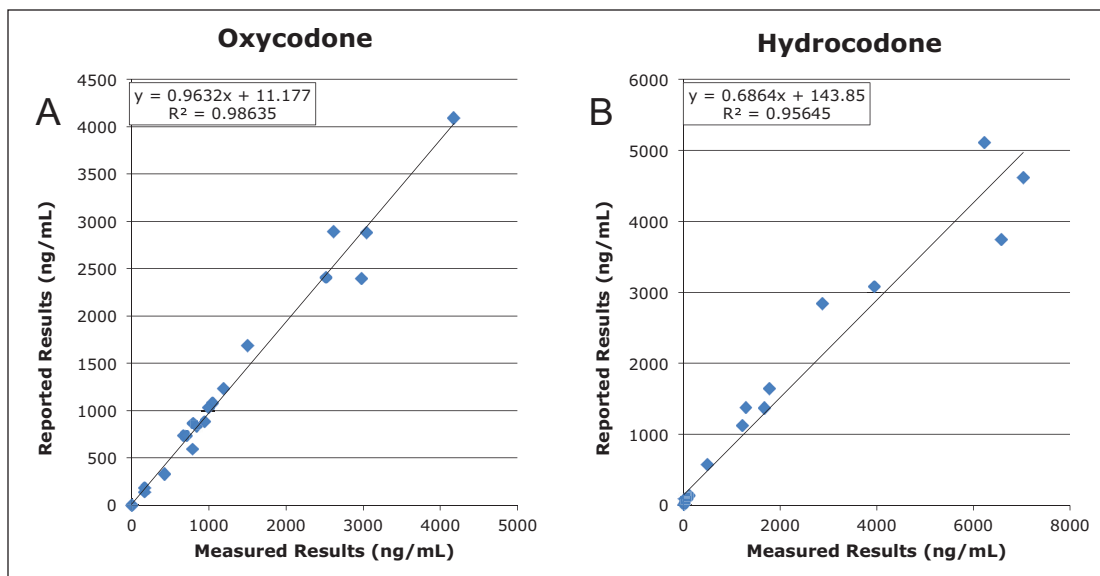


Figure 4. Comparison of results obtained using the current method vs. an alternative LC/MS/MS confirmation method for previously analyzed incurred samples.

A significant difference was seen when the samples were analyzed for compounds such as morphine, oxycodone, and hydromorphone that undergo significant glucuronidation prior to excretion. Many methods used to analyze opioid drugs rely on enzymatic hydrolysis. However, the degree of hydrolysis is greatly dependent upon not only the  $\beta$ -glucuronidase enzyme used (ex: *Patella vulgate*, *Helix pomata*, *Escherichia coli*), but also on the substrate (morphine-6-gluc vs. morphine-3-gluc, morphine-3-gluc vs. hydromorphone-3-gluc).<sup>6</sup> Analysis of the same group of samples by the current and previously reported methods revealed that the reliance on enzymatic hydrolysis dramatically underestimates the total amount of glucuronidated metabolites. Regression analysis of reported released oxycodone and hydromorphone vs. the actual measured totals of each compound using the current method (glucuronide conjugate + free drug) yielded slopes of 0.20 and 0.25, respectively, indicating that 75% to 80% of the drug was not hydrolyzed. Analysis with this current method reveals that > 85% of total oxycodone and hydromorphone exist as glucuronide conjugates. Thus, any inefficiencies in glucuronide hydrolysis could result in significant underestimation of total compound concentration. The current method, obviously, is not subject to this limitation, since glucuronide metabolites are measured directly.

## CONCLUSIONS

The method presented here demonstrates the advantages of mixed-mode  $\mu$ Elution SPE combined with UPLC/MS/MS for the analysis of 26 opioid compounds and metabolites of interest. All compounds were analyzed in under 5.5 min with complete resolution of all isobaric compound pairs. The use of Oasis MCX  $\mu$ Elution Plates resulted in improved linearity, and significantly reduced matrix effects compared to a simple dilution method. Accuracy and precision for quality control samples and calibration standards were also improved using mixed-mode SPE. The ability to achieve LOQs of 5 ng/mL for nearly all analytes and the ability to measure glucuronide metabolites directly without hydrolysis make this method well suited for the analysis of these compounds.

## References

1. Goldberger, B.A. and E.J. Cone, Confirmatory tests for drugs in the workplace by gas chromatography-mass spectrometry. *Journal of Chromatography A*, 1994. 674(1-2): p. 73-86.
2. Gustavsson, E., et al., Validation of direct injection electrospray LC-MS/MS for confirmation of opiates in urine drug testing. *Journal of Mass Spectrometry*, 2007. 42(7): p. 881-889.
3. Murphy, C.M. and M.A. Huestis, LC-ESI-MS/MS analysis for the quantification of morphine, codeine, morphine-3- $\beta$ -D-glucuronide, morphine-6- $\beta$ -D-glucuronide, and codeine-6- $\beta$ -D-glucuronide in human urine. *Journal of Mass Spectrometry*, 2005. 40(11): p. 1412-1416.
4. Edinboro, L.E., R.C. Backer, and A. Poklis, Direct Analysis of Opiates in Urine by Liquid Chromatography-Tandem Mass Spectrometry. *Journal of Analytical Toxicology*, 2005. 29(7): p. 704-710.
5. French, D., A. Wu, and K. Lynch, Hydrophilic interaction LC-MS/MS analysis of opioids in urine: significance of glucuronide metabolites. *Bioanalysis*, 2011. 3(23): p. 2603-2612.
6. Wang, P., et al., Incomplete Recovery of Prescription Opioids in Urine using Enzymatic Hydrolysis of Glucuronide Metabolites. *Journal of Analytical Toxicology*, 2006. 30(8): p. 570-575.
7. Watts, R., et al., Simultaneous quantitative determination of opioid dependency treatment drugs in human urine using UPLC-MS/MS. Waters Application Note, 2012.
8. Smith, H.S., Opioid Metabolism. Mayo Clinic proceedings. Mayo Clinic, 2009. 84(7): p. 613-624.

**This information is provided for clinical research purposes and is not intended for clinical diagnostic use.**

# Waters

THE SCIENCE OF WHAT'S POSSIBLE.™

Waters, Oasis, ACQUITY UPLC, Xevo, and UPLC are registered trademarks of Waters Corporation. MassLynx and The Science of What's Possible are trademarks of Waters Corporation. MilliQ is a registered trademark of Millipore Corporation. Cerilliant is a registered trademark of Cerilliant Corporation. All other trademarks are the property of their respective owners.

©2013 Waters Corporation. Produced in the U.S.A.  
April 2013 720004650EN AG-PDF

**Waters Corporation**  
34 Maple Street  
Milford, MA 01757 U.S.A.  
T: 1 508 478 2000  
F: 1 508 872 1990  
[www.waters.com](http://www.waters.com)

